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DESIGNING OF GLUTAMATE RECEPTOR INHIBITORS OF QUINAZOLINONE DERIVATIVES BY A COMPARATIVE QSAR ANALYSIS AND MOLECULAR MODELING STUDIES

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ABSTRACT

An attempt was made to develop a two dimensional quantitative structure-activity relationship (2D-QSAR) and molecular docking studies on a series of quinazolinone derivatives acting as glutamate receptor inhibitors for correlating the chemical composition of quinazolinone analogs and estimation of their anticonvulsant activity using Multiple Linear Regression (MLR) Analysis. New Chemical Entities (NCEs) were designed using results of pharmacophore profiling from known anticonvulsants. Binding affinities of designed NCEs were studied on Glutamate receptor using docking studies and their ADMET properties were also predicted. Finally, most promising compounds were selected from molecular modeling studies. 12 compounds showed significant glutamate receptor inhibiting activity compared to standard ligand bound with glutamate receptor (PDB: 1GR2). These four basic strategies (Pharmacophore mapping, QSAR, docking and ADMET screening) were implemented to evaluate the performance of derivatives. Although predicted Ki through QSAR model showed mild activity against glutamate receptor, but conclusively, compounds 22, 15 and 8 were observed to be most feasible to act against glutamate receptor for anticonvulsant activity.

KEYWORDS: ADMET, Anticonvulsant, Docking, Glutamate, Pharmacophore, QSAR & Quinazolinone

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INTRODUCTION

Epilepsy, a neurological, non-communicable, pervasive disease signalized by the unpredictable and periodic occurrence of a transient alteration of behavior due to the disordered, synchronous and rhythmic firing of populations of brain neurons that affects more than 60 million people worldwide according to epidemiological studies ^{1,2}. In developed countries approximately 50 per 100,000 while that of developing country is 100 per 100,000, in 2013 about 22 million people have been suffering from epilepsy (WHO, 2006). The presently available antiepileptic drugs are unable to control seizures effectively in as many as 25% of the patients³. Many conventional antiepileptic drugs like phenytoin, sodium valproate and carbamazepine reported several serious side effects notably neurotoxicity with them ^{4,5}. The greater numbers of antiepileptic drugs are consumed life long, concomitant administration of other drugs predisposes to the risk of drug interaction. However, newer antiepileptics like gabapentin, vigabatrin, lamotrigine, *etc* are used supplemental to the conventional agents⁶.

Anticonvulsant drugs are estimated to be effective in treating 90% of the epileptic patients only. However, all the conventional and newer anticonvulsant drugs which are currently approved, and are already in use, have dose-related toxicity and idiosyncratic side effects⁷. Thus, it is essential to investigate for new

antiepileptic agents with lower toxicity and fewer side effects. A wide variety of compounds has been designed for this purpose⁸.

The reason behind epilepsy has been defined on the basis of single gene defect, interaction of multiple genes as well as an environmental factor⁹. Most of the interacting genes are known to be involved with ion channels, enzymes, gamma-amino-butyric-acid (GABA), and G protein-coupled receptors ¹⁰. The majority of antiepileptic drugs reduce the release of excitatory glutamate by blocking sodium or calcium channels, activation of GABA, inhibition of glutamate receptor and activation of peroxisome proliferator-activated receptor alpha. At present, the most commonly used anticonvulsant therapy for synaptic transmission are through the concomitant use of drugs that belongs either to the class of GABA activator (GA), Na^{+/}Ca⁺⁺ inhibitor (NCaI), glutamate receptor inhibitor (GRI), and PPAR-alpha activator (PPARα) *etc.* ^{11,12}.

Synaptic transmission in the mammalian central nervous system is mediated by 'L-glutamate' on three classes of ionotropic glutamate receptors, namely AMPA, NMDA and Kainate. GRIs work through these receptors. GRI (Glutamate receptor inhibitor) are structurally diverse group of compounds which binds to the glutamate receptor and interacts with a specific allosteric non-substrate binding pocket site. Currently, drugs used to treat epilepsy under GRI for anticonvulsant therapy are Perampanel ^{13, 14}.

As quinazolinone backbone has shown a variety of biological activities it appears as an ideal frame for designing of anti-epileptic leads as glutamate receptor inhibitor. They may act as anticonvulsant potential lead as well as possess efficacy against epilepsy. Crystal structure analysis of glutamate receptor (PDB: 1GR2, 4PE5) showed that most of GRI bound to the glutamate receptor. One of the wings of this GRI interact with a hydrophobic pockets formed mainly by the side chains of tyrosine, phenylalanine, threonine, arginine, serine, glutamic acid. Recently, pharmacophore mapping, 2D/3D QSAR and docking guided optimization of identification of novel compounds have been used as important strategies in the discovery of new anticonvulsants. ADMET screening of newly designed molecules provide a pre-clinical trial scaled analysis for their bioavailability and drug-like possibilities. These approaches are inexpensive and more practical than discovering novel compounds. The present work was focused on computer-aided design of GRI containing quinazolinone nucleus with simultaneous goals of enhanced performance against glutamate receptor. All the New Chemical Entities (NCEs) were designed on the basis of pharmacophore components of well known anticonvulsants from literature survey. In order to gain molecular interaction insights, docking studies of NCEs are carried out targeting glutamate receptor. The possible activity of NCEs can be obtained from two-dimensional (2D) quantitative structureactivity relationship (QSAR) studies using multiple linear regression (MLR) Analysis. ADMET properties are used to estimate the drug like pharmacokinetic profile of the designed NCEs. The most promising compounds can be selected on the basis of results of molecular modeling studies. After confirmation of molecular interaction, their activity and ADMET screening of derivatives were performed for cross-evaluation of their performance for identification of most possible novel compounds 15-20.

MATERIAL AND METHODS

Raw Data

Raw data for anticonvulsant activity was collected from literatures and databases. Anticonvulsant activity was collected in terms of Ki (nM) value against glutamate receptor. Bioactivity in term of Ki nM (inhibition constant) was transformed to (natural) log Ki (nM) for normalisation of data set. Crystallographic structure of glutamate receptor was

collected from PDB database.

Pharmacophore Profiling of Compounds

Pharmacophore properties of anticonvulsant compounds were identified through literatures. These properties were used for designing of quinazolinone derivatives. The pharmacophore profiles of designed molecules were judged in comparison of positive control compounds.

Structural Modelling and Optimization

Chem Bio Draw Ultra v12.0 modeling suite (CambridgeSoft Corp., UK) was used for sketching of compounds under study. Molecule's geometry cleaning and energy minimization was performed by Discovery studio 3.5 client (Accelrys USA). It was also used for conversion of 2D to 3D structure.

Docking Simulation Parameters

Auto Dock Vina 4.2 was used for virtual high throughput screening of compounds against glutamate receptor. Docking of known positive control was used for identification of best possible binding site of query molecules. During docking simulation process, ligand was set to flexible mode, while the protein set to rigid form. All other docking simulation parameters were set to default mode²¹.

Chemical Descriptors and QSAR Modelling Parameters

To screen out potential leads against glutamate receptor, a total of known anticonvulsant compounds with low to high Ki (nM) values were collected in the raw data set from PubChem database of NCBI^{22.} To select the compounds for model development, pharmacophore features of control and query compounds were matched. Only best selected compounds were used for model building. Molecular descriptors were calculated through PaDEL-Descriptors software²³. After removing zero values descriptors, the descriptors were selected through data reduction through removal of highly inter-correlated descriptors followed by forward selection and backward elimination procedures. Finally, a total of 17 known anticonvulsant compounds with experimental Ki and two molecular descriptors were found to be involved in the model building using multiple-linear-regression (MLR) method. QSAR model robustness and prediction quality were represented by high regression coefficient (R²) value. Cross-validation of QSAR model was done by LOO (Leave-one-out) approach. The applicability domain of derived QSAR model was indicated by cross-validation regression coefficient (R²_{CV}). Evaluation of model was also performed through residual plot.

Evaluation of Pharmacokinetic Behaviour through Lipinski's Rule of Five and ADME Parameters

Potential leads may fail to clear the clinical trial approval through FDA due to unmatched standard pharmacokinetic properties. The key pharmacokinetic properties were represented by 'admetSAR' as used by Drug Bank database²⁴. Besides this, Lipinski's rules of five²⁵ along with other physicochemical properties were used to explain the pharmacokinetic behaviour of compounds. Topological polar surface area (TPSA) and molecular weight (MW) (cutoff= \leq 500) were used to evaluate the fractional absorption of compounds. Bioavailability of compounds were evaluated by topological PSA (polar surface area) (cutoff= \leq 140 Å²). These descriptors also represent the passive membrane transport. For estimation of fractional absorption, sum of H-bond donors and acceptors was used. Additionally, number of rotatable bonds also used as a measure of bioavailability. The pharmacokinetic behaviour of drug distribution depends on membrane permeability (estimated by Caco-2 cell line), blood-brain barrier and distribution (volume). Excretion ability of

compounds from the body is evaluated on the basis of logP (octanol/water) and molecular weight. Renal clearance is indicated by negative lipophilicity of molecule. Metabolism of compounds in liver was evaluated on the basis of logP value (hydrophobic condition) and topological polar surface area of molecules. Lipophilicity of molecule also provided indications about absorption and metabolic process. Majority of oral bio-available drugs (90%) follow the Lipinski's rule of five; therefore the designed molecules were also studied for oral bioavailability of active anticonvulsant drugs through rule of five. These chemical properties for drug-likeness were calculated for quinazolinone derivatives and further evaluated for compliance with a standard drug.

RESULTS AND DISCUSSIONS

Preliminary quantitative structure-activity relationship (QSAR) studies revealed high structure-activity relationship (*i.e.* pharmacophore) features for anticonvulsant activity. Based on features identified from pharmacophore, molecules were designed on the nucleus of quinazolinone. Pharmacophore features were also used for data collection for QSAR model building using multiple linear regression (MLR) method. Since the QSAR approach is a well established as lead optimization method, therefore the designed molecules were screened through QSAR model to predict the Ki value of new anticonvulsant compounds derivatives, therefore indicating the activity range. The binding affinity of known anticonvulsant target glutamate receptor was studied through docking simulation so that to identify the possible binding site and to explain the drug-target activity relationship by using the crystallographic complex structure of glutamate receptor. Finally the designed molecules were processed for ADMET screening for estimate the pharmacological behaviour of designed molecules. Results of pharmacophore, QSAR based Ki prediction, docking, and ADMET screening were analysed to receive conclusive information to predict the possibilities about designed molecules for anticonvulsant activity.

Pharmacophore Profiling for Anticonvulsant Activity through Glutamate Receptor Inhibition

The known quantitative structure-activity-relationship studies revealed possible pharmacophore features for anticonvulsant activity. The designed compounds possess the pharmacophore essential for anticonvulsant activity. The pharmacophore proposed contains: (i) hydrophobic domain (HPD) of newer anticonvulsants considering the most potent of semicarbazones as anticonvulsant recently, it was planned to prepare new semicarbazones with quinazoline scaffold; (ii) hydrogen bonding donors (HBD) (iii) two electron donor system (D); and (iv) distal aryl ring which affects pharmacokinetics (PKS). All these elements are present in other clinically effective drugs or their metabolites.

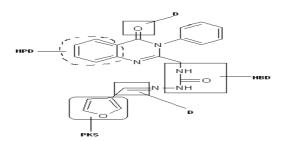


Figure 1: Example showing Pharmacophore components at quinazolinone derivative

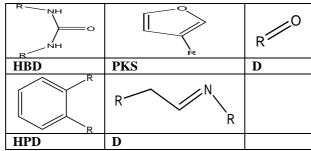


Figure 2: Pharmacophore components used for designing of quinazolinone derivatives

Pharmacophore components in quinazolinone derivatiove are as: HBD (Hydrogen Bonding Domain), PKS (A diastral aryl ring which affects pharmacokinetics), HPD (Hydrophobic domain), D (Electron donor system)

To understand the pharmacophore behavior of glutamate inhibitor for anticonvulsant activity, model building molecules were observed. Observations were based on the sub-structural relationships considering the electron flow. It was found that the following pharmacophore components are available for anticonvulsant activity through inhibition of glutamate receptor: hetero-non-aromatic 5-point ring, an amine group, a hydroxyl group, imine group, carboxylic acid group, and benzene ring. Our designed molecules did not contain these pharmacophore components. This observation also represents the comparatively mild potential of designed molecules in comparison of existing inhibitors of glutamate receptor.

Design of New Chemical Entities (NCEs) Containing Quinazolinone Nucleus

The findings of pharmacophore studies provided the overall substitution pattern (electrostatic, steric and hydrophobic pattern) required around the quinazolinone nucleus. Hypotheses shown in literature were also considered for optimization of quinazolinone derivatives as shown in figure 3. Pharmacophore features signified the importance of quinazolinone nucleus for the anticonvulsant activity of compounds. This information had helped a lot in optimizing quinazolinone pharmacophore and designing of NCEs containing quinazolinone ring for potent anticonvulsant activity. Substitution pattern around quinazolinone pharmacophore showed in Figure 2 was used for the manual design of NCEs. Designed compounds were passed through Lipinski's screen to ensure drug like the pharmacokinetic profile of the designed compounds in order to improve their bioavailability. The parameters used as Lipinski's filters are: Number of hydrogen bond acceptor (A) (<10), number of hydrogen bond donor (B) (<5), number of rotatable bond (R) (<10), Clog P (X) (<5), molecular weight (W) (<500 g/ mol) and polar surface area (S) is (<140 Å).

We had designed twenty-one compounds containing quinazolinone nucleus with substitution pattern shown in figure 4. All these compounds were subjected for further studies to sort out the compounds with good binding affinity for glutamate receptor and having good ADME properties.

$$R_2$$

Figure 3: Scaffold Used in Designing of Quinazolinone Derivatives

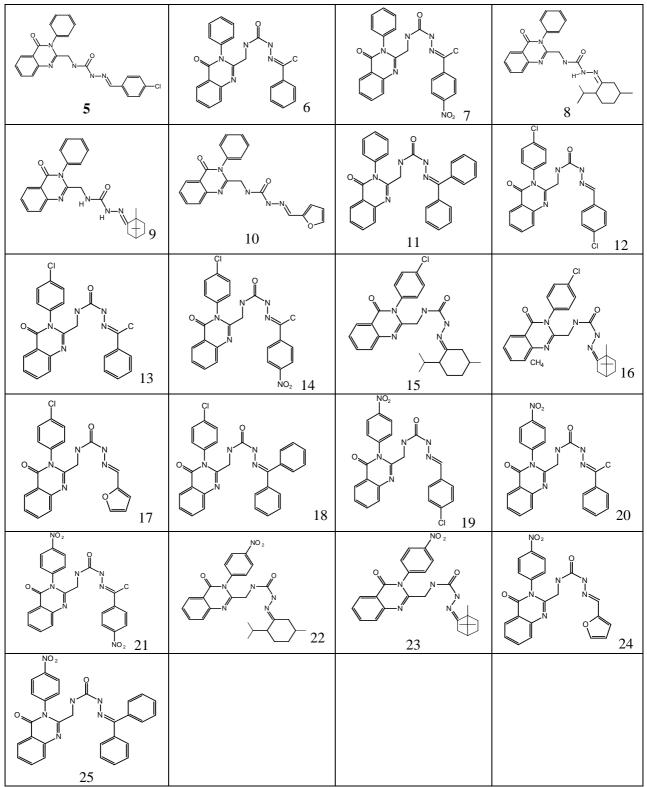


Figure4: Designed Quinazolinone Derivative

Molecular Docking Studies on Glutamate Receptor

To understand the binding behavior of studied compounds named '5 to 25' against glutamate receptor-based anticonvulsant activity were processed for docking studies. The docking studies suggested that designed compounds inhibit the glutamate receptor activity by high binding affinity in comparison to control. Later, orientations and binding affinity of

tested derivatives were explored. The binding affinity allowed the derivatives to be compared with standard control 'Kainate'.

Docking Studies

Molecular docking tool, AutoDock-Vina software was used for studying binding modes of the designed compounds into the binding pocket of glutamate receptor. AutoDock-Vina was found to produce the least number of inaccurate poses and results near to native co-crystallized structures ²⁶. These studies helped to sort out the designed compounds with good binding affinity against glutamate receptor. The docking score in terms of Kcal/mol and other results of docking studies of designed compounds of quinazolinone series are presented in Table 1.

Binding Affinity

Binding affinity is shown in a negative value, which indicates the measure of the stability of ligand-Protein interaction. The binding affinity of the standard compound Kainate (PDB: 1GR2) was found to be -8.2 kcal/mol. The binding affinity of the designed NCEs "10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 5, 6, 7, 8, 9" was found to be -8.5, -9.7, -8.6, -8.4, -9.4, -9, -8.4, -8.3, -9.1, -8.3, -8.7, -9.2, -8.8, -8.7, -8.6, -9.3, -8.9, -9.1, -9.4, -9.1, and -9 kcal/mol respectively. The close analysis of these results suggests that the designed NCEs have a comparable binding affinity with the standard compound. Overall interacting residues can be visualized from Table-1.

Contacts

Docking studies were analyzed in reference to known target (Glutamate receptor) binding ligand control IRG2. Literature based information shows that binding pocket of control ligand at target bears tyrosine, glutamic acid, proline, threonine, arginine, and serine residues majority of hydrophobic nature. Control ligand binds with a binding affinity of -8.1 kcal/mol. In reference of control ligand, query compound shared common residues tyrosine, glutamic acid, proline, threonine, arginine, and serine. Out of the tested ligand compounds, "10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 5, 6, 7, 8, 9" ligands were found to be possibly good interactivity with target Glutamate receptor.

Table 1: Docking Results and Evaluation of Query Molecules in Reference of Known Glutamate Receptor Binder at PDB: 1GR2

| | | Binding Affinity (kcal/mol) | Interacting Residues | Comparison with Control |
|---------|---------------------------|-----------------------------------|--|-------------------------|
| Control | Kainate (PDB: 1GR2 bound) | -8.1 | Tyrosine, Glutamic Acid, Proline, Threonine, Arginine, And Serine | |
| Query | 10 | -8.5 | Arginine, Glutamic Acid | Stronger |
| Query | 11 | -9.7 | Arginine, Glutamic Acid | Stronger |
| Query | 12 | -8.6 | Arginine, Serine | Stronger |
| Query | 13 | -8.4 | Tyrosine (Pi-Pi Interaction), Arginine | Stronger |
| Query | 14 | -9.4 | Tyrosine (Pi-Pi Interaction), Srginine, Serine, Threonine | Stronger |
| Query | 15 | -9 | Tyrosine (Pi-Pi Intercation), Glutamic Aicd | Stronger |
| Query | 16 | -8.4 | Tyrosine, Glutamic Acid | Stronger |
| Query | 17 | -8.3 | Arginine, Serine | Stronger |
| Query | 18 | -9.1 | Tyrosine (Pi-Pi Intercation), Glutamic Acid | Stronger |
| Query | 19 | -8.3 | Tyrosine (Pi-Pi Intercation), Glutamic Acid | Stronger |

| Query | 20 | -8.7 | Arginine (Sigma-Pi Bond), Glutamic Acid | Stronger |
|-------|----|------|--|----------|
| Query | 21 | -9.2 | Tyrosine (Pi-Bond), Serine, Arginine, Threonine | Stronger |
| Query | 22 | -8.8 | Tyrosine (Pi-Bond) | Stronger |
| Query | 23 | -8.7 | Tyrosine (Pi-Bond), Glutamic Acid | Stronger |
| Query | 24 | -8.6 | Arginine, Glutamic Acid, Serine | Stronger |
| Query | 25 | -9.3 | Tyrosine, Threonine, Glutamic Acid | Stronger |
| Query | 5 | -8.9 | Arginine, Glutamic Acid | Stronger |
| Query | 6 | -9.1 | Arginine, Glutamic Acid | Stronger |
| Query | 7 | -9.4 | Tyrosine (Pi-Pi Interaction), Serine, Arginine, Threonine | Stronger |
| Query | 8 | -9.1 | Arginine, Glutamic Acid | Stronger |
| Query | 9 | -9 | Threonine, Glutamic Acid | Stronger |

Table 1.1: Residual Interaction with '5 to 25'

| Interacting Residue | ID | Residual Energy (Kcal/Mol) |
|----------------------------|-----|----------------------------|
| ALA | 75 | -2.3718 |
| ARG | 108 | -3.71365 |
| ARG | 188 | -4.52691 |
| ASN | 84 | -1.20303 |
| ASP | 70 | -0.45587 |
| ASP | 155 | -0.40623 |
| GLU | 25 | -6.10361 |
| GLY | 71 | -6.37509 |
| GLY | 74 | -6.32285 |
| GLY | 157 | -5.48117 |
| ILE | 23 | -1.61984 |
| LEU | 24 | -0.58823 |
| LEU | 154 | -7.82805 |
| LYS | 72 | -14.0569 |
| MET | 212 | -0.53573 |
| SER | 156 | -11.4711 |
| SER | 158 | -0.58833 |
| THR | 189 | -12.3745 |
| THR | 190 | -7.53074 |
| TYR | 73 | -25.9379 |

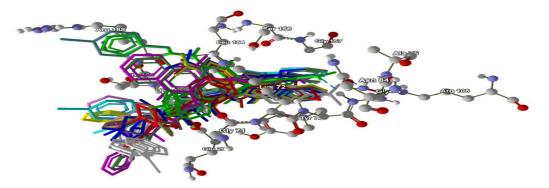


Figure 5 A: Residual interaction of glutamate receptor with '5 to 25' & Docking results of quinazolinone derivatives against glutamate receptor

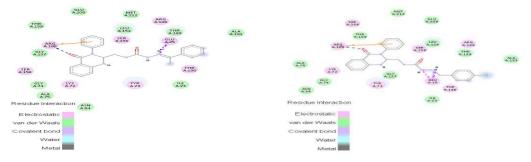


Figure 5.A: 5 at Glutamate Receptor PDB: 1GR2 Figure 5.B: 6 at Glutamate Receptor (PDB: 1GR2)



Figure 5.C: 7 at Glutamate Receptor (PDB: 1GR2 Figure 5.D: 8 at Glutamate Receptor (PDB: 1GR2)



Figure 5.E: 9 at glutamate receptor (PDB: 1GR2) Figure 5.F: 10 at glutamate receptor (PDB: 1GR2)

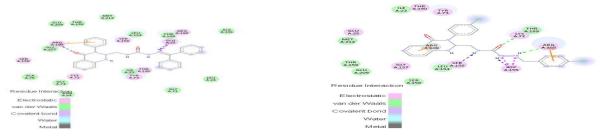


Figure 5.G: 11 at Glutamate Receptor (PDB: 1GR2) Figure 5.H: 12 at Glutamate Receptor (PDB: 1GR2)

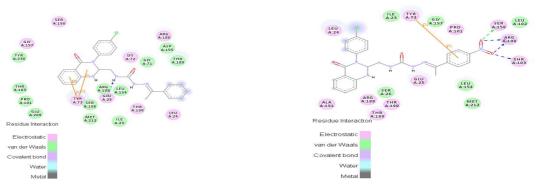


Figure 5.I: 13 at Glutamate Receptor (PDB: 1GR2) Figure 5.J: 14 at Glutamate Receptor (PDB: 1GR2)

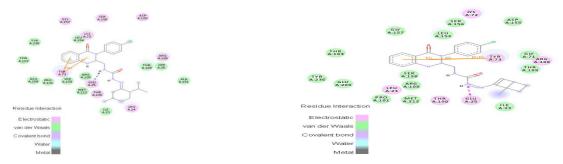


Figure 5.K: 15 at Glutamate Receptor (PDB: 1GR2) Figure 5.L: 16 at Glutamate Receptor (PDB: 1GR2)

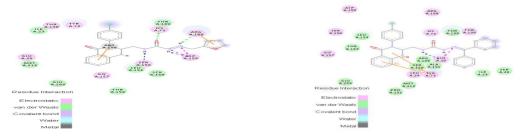


Figure 5.M: 17 at Glutamate Receptor (PDB: 1GR2)

Figure 5.N: 18 at Glutamate Receptor (PDB: 1GR2)

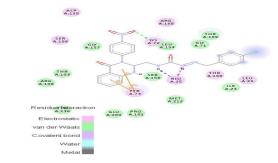


Figure 5.O: 19 at Glutamate Receptor (PDB: 1GR2)

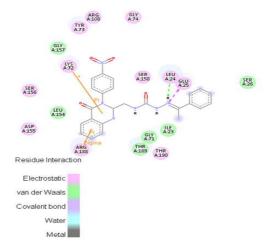


Figure 5.P: 20 at Glutamate Receptor (PDB: 1GR2)

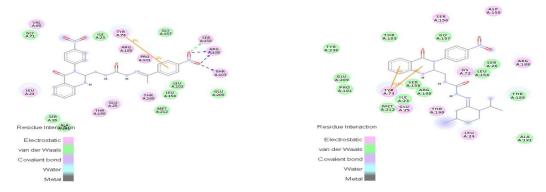


Figure 5.Q: 21 at Glutamate Receptor (PDB: 1GR2) Figure 5.R: 22 at Glutamate Receptor (PDB: 1GR2)

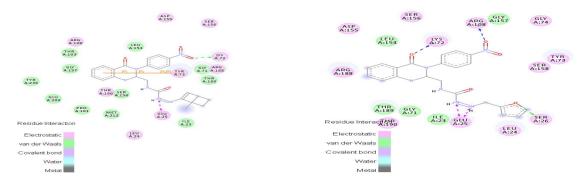


Figure 5.S: 23 at Glutamate Receptor (PDB: 1GR2) Figure 5.T: 24 at Glutamate Receptor (PDB: 1GR2)

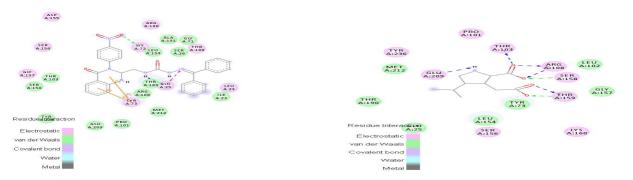


Figure 5.U: 25 at Glutamate Receptor (PDB: 1GR2) Figure 5.V: Kainate at Glutamate Receptor (PDB: 1GR2)

Pharmacophore Distance Map

Structure-based pharmacophore distance map was prepared for quinazolinone derivatives, where were found to be mild active against glutamate receptor and GABA respectively. Distance map was prepared on the basis of six properties from the designed molecules. The properties were as: hydrogen bond acceptor, hydrogen bond donor, hydrophobic, negative ionizable component, positive ionizable and ring aromatic. The maps were drawn through manual process by measuring the average inter-component distance from the designed molecules. The designed glutamate receptor binders showed only five-property based pharmacophore, as shown in following figure 6.

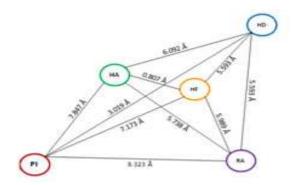


Figure 6: Five property based Pharmacophore distance map of Quinazolinone designed molecules against Glutamate receptor. Here the components are as: Hydrogen Bond Acceptor (HA), Hydrogen Bond Donor (HD), Hydrophobic (HF), Positive Ionisable (PI) and Ring Aromatic (RA) components.

The inter-component distance has been providing in Å.

QSAR Model Based Prediction of Glutamate Targeted Anticonvulsant Activity QSAR studies

All QSAR studies were performed in Weka software. A series of 22 compounds of quinazolinone derivatives tested for their anticonvulsant activity was selected for QSAR Studies. 17 Compounds were used for model building. The model was cross evaluated with Leave-One-Out-Cross validation (LOOCV) method. Selection of molecules in the training set and their cross validation is a key and important feature of any QSAR model. Therefore the care was taken in such a way that biological activities of all compounds validation result must *lie* within the maximum and minimum value range of biological activities of the training set of compounds. A Uni-column statistics for the training set and the validated result were generated to check the correctness of selection criteria for training set molecules. The maximum and minimum value in training and set were compared in a way that:

- The maximum value of LN Ki (nM) of query compound should be less than or equal to the maximum value of LN Ki (nM) of the training set.
- The minimum value of LN Ki (nM) of query compound should be higher than or equal to the minimum value of LN Ki (nM) of the training set.

This observation showed that validation was interpolative and derived within the minimum–maximum range of training set. The mean and standard deviation of LN Ki (nM) values of sets of training and test provide insights to the relative difference of mean and point density distribution of two sets. Several 2D QSAR models were generated for a training set of 17 compounds using MLR method. The best QSAR model was selected on the basis of the value of statistical parameters like R^2 (square of the correlation coefficient for a training set of compounds), and R^2_{cv} (LOO cross-validated R^2). The QSAR model was validated through LOOCV method. Statistical results generated by 2D QSAR analysis showed that QSAR model has good cross-validation predictability. Prior studies of anticonvulsant lead identification and optimization showed an important part of QSAR application in drug discovery. Activity was predicted on the basis of derived QSAR model for newly designed quinazolinone derivatives. The QSAR model development accuracy was represented by R^2 (= 0.924) (i.e., 92.4%) and activity prediction accuracy denoted by R^2_{CV} (= 0.889) (i.e., 88.9%) (Table 2, 3) (Figure 7, 8). Two chemical descriptors namely, ATSc4 and VCH.7 well allied with experimental anticonvulsant activity. Derived QSAR model equation as:

Predicted LN Ki (nM) = 6.7103 * D2 + 9.2198 * D3 + 5.4257

(Here D2: ATSc4, D3: VCH.7; these are molecular descriptors calculated from PaDEL-Descriptor software)

$$[R^2 = 0.924 \text{ and } R^2_{CV} = 0.889]$$

Where, R^2 = regression coefficient and R^2_{CV} = cross-validation regression coefficient. QSAR results suggest that compounds '8, 15 and 22' possess good potency for anticonvulsant activity.

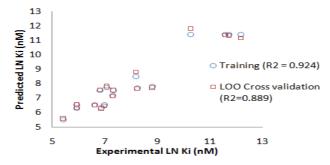


Figure 7: Regression Plots showing model training as well as cross-Evaluation results through Leave-One-Out Cross Validation

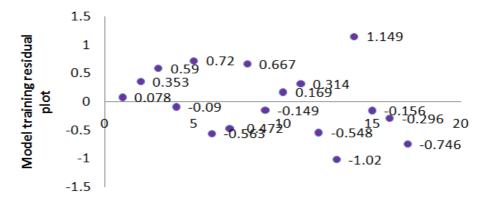


Figure 8: Residual Plot of Model Representing Error Distribution of QSAR Model

Table 2: Model Development & LOO Cross Validation

| Dataset | CHEMBL ID | D2 | D3 | Experimental LN_Ki (nM) | Training predicted | Training Error | CV LOO Predicted | CV LOO Error |
|---------|--------------|----------|----------|----------------------------|--------------------|-------------------|---------------------|-----------------|
| Train | 106579 | -0.10068 | 0.083036 | 5.438079 | 5.516 | 0.078 | 5.578 | 0.14 |
| Train | 110915 | 0.015835 | 0.082566 | 5.940171 | 6.293 | 0.353 | 6.596 | 0.656 |
| Train | 317184 | 0.044173 | 0.087679 | 5.940171 | 6.531 | 0.59 | 6.347 | 0.407 |
| Train | 110866 | 0.044173 | 0.087679 | 6.620073 | 6.531 | -0.09 | 6.521 | -0.099 |
| Train | 286782 | 0.09555 | 0.160209 | 6.824374 | 7.544 | 0.72 | 7.596 | 0.772 |
| Train | 107990 | 0.044605 | 0.064998 | 6.887553 | 6.324 | -0.563 | 6.256 | -0.632 |
| Train | 323142 | 0.044186 | 0.087679 | 7.003065 | 6.531 | -0.472 | 6.478 | -0.525 |
| Train | 280179 | 0.124835 | 0.160209 | 7.07327 | 7.74 | 0.667 | 7.804 | 0.731 |
| Train | 110618 | 0.138566 | 0.087679 | 7.31322 | 7.164 | -0.149 | 7.142 | -0.171 |
| Train | 415462 | 0.092181 | 0.160209 | 7.352441 | 7.521 | 0.169 | 7.533 | 0.181 |
| Train | 432781 | 0.274668 | 0.133869 | 8.188689 | 8.503 | 0.314 | 8.816 | 0.628 |
| Train | 25875 | 0.11994 | 0.160209 | 8.255828 | 7.708 | -0.548 | 7.658 | -0.597 |
| Train | 23255 | 0.134464 | 0.160209 | 8.824678 | 7.805 | -1.02 | 7.695 | -1.13 |
| Train | 2115153 | 0.088826 | 0.585974 | 10.27505 | 11.424 | 1.149 | 11.8 | 1.525 |
| Train | 2114116 | 0.088826 | 0.585974 | 11.58058 | 11.424 | -0.156 | 11.373 | -0.207 |

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| Train | 2115156 | 0.088826 | 0.585974 | 11.71994 | 11.424 | -0.296 | 11.328 | -0.392 |
|-------|---------|----------|----------|----------|--------|--------|--------|--------|
| Train | 2115151 | 0.088826 | 0.585974 | 12.17045 | 11.424 | -0.746 | 11.18 | -0.99 |

Table 3: Query Prediction through QSAR Model

| Data Set | ID | D2 | D3 | Model Predicted LN Ki (nM) | Z-Score | Calculated Ki (µM) |
|-------------|---|----------|----------|-------------------------------|----------|-----------------------|
| Query | Kainate (Exp. Ki: 477 to 12221 nM) PDB:1GR2_Glutamate_Control | 0.168399 | 0.200681 | 8.406 | 0.152137 | 4.473829 |
| Query | 9 | 0.09199 | 0.134052 | 7.279 | -0.37837 | 1.449538 |
| Query | 22 | 0.156003 | 0.200681 | 8.323 | 0.113067 | 4.117494 |
| Query | 19 | 0.06611 | 0.200681 | 7.72 | -0.17078 | 2.25296 |
| Query | 20 | 0.071504 | 0.200681 | 7.756 | -0.15383 | 2.335544 |
| Query | 21 | 0.070689 | 0.200681 | 7.75 | -0.15666 | 2.321572 |
| Query | 7 | 0.083085 | 0.200681 | 7.833 | -0.11759 | 2.522485 |
| Query | 5 | 0.078506 | 0.200681 | 7.803 | -0.13171 | 2.447935 |
| Query | 18 | 0.07172 | 0.200681 | 7.757 | -0.15336 | 2.33788 |
| Query | 6 | -0.00301 | 0.200681 | 7.256 | -0.3892 | 1.416579 |
| Query | 25 | 0.084116 | 0.200681 | 7.84 | -0.11429 | 2.540205 |
| Query | 23 | 0.071059 | 0.200681 | 7.753 | -0.15525 | 2.328548 |
| Query | 13 | 0.07881 | 0.200681 | 7.805 | -0.13077 | 2.452836 |
| Query | 14 | 0.077996 | 0.200681 | 7.799 | -0.13359 | 2.438163 |
| Query | 15 | 0.16331 | 0.200681 | 8.372 | 0.136132 | 4.324276 |
| Query | 12 | 0.073417 | 0.200681 | 7.769 | -0.14771 | 2.366104 |
| Query | 17 | 0.093333 | 0.200681 | 7.902 | -0.08511 | 2.702682 |
| Query | 11 | 0.079027 | 0.200681 | 7.806 | -0.1303 | 2.45529 |
| Query | 24 | 0.105728 | 0.200681 | 7.985 | -0.04604 | 2.936577 |
| Query | 16 | 0.078366 | 0.200681 | 7.802 | -0.13218 | 2.445488 |
| Query | 10 | 0.100639 | 0.200681 | 7.951 | -0.06204 | 2.838412 |
| Query | 8 | 0.164106 | 0.203203 | 8.4 | 0.149313 | 4.447067 |

Compliance with Pharmacokinetics Properties and Toxicity Estimation (ADME/T)

Prediction of the ADME parameters prior to the experimental studies is one of the most important aspects of the drug discovery and development of the drug molecule. The drug may fail to reach the market phase if those properties are not fulfilled by the drug candidate. Taking into consideration the above-mentioned aspects, the ADME profile of the designed NCEs was studied using the tool admeSAR. In addition to predicting molecular properties, provides the ranges for comparing the properties of the molecules with those of majority of known drugs. The range of values that cause a molecule to be flagged can be similar or dissimilar to other known drugs. Lipinski's rule of five and adme-SAR physical descriptors and pharmaceutically relevant properties of quinazolinone analogs were analyzed, among which significant descriptors were reported here required for predicting the drug-like properties of molecules. These properties were (Table 4)

Glutamate receptor's control inhibitors were observed to bear the properties of crossing the Blood-brain-Barrier (BBB), being intestinal absorbable, easy accessibility to the cells, the non-inhibitor substrate of plasma proteins, being safe from being metabolized through CYPs and don't have carcinogenic properties. Results revealed that quinazolinone derivative (5 to 25) followed the screening through Lipinski's rule of five for oral bioavailability, while 18 and 25 showed an acceptable violation of 1 and 2. Here, the hydrophilicity of studied compounds was measured by logP value. Rule of five screening results indicate mild hydrophilicity of quinazolinone derivatives and so there is the good average possibility

of absorption or membrane permeability due to their logP values less than 5 (Table 4). LogP also associated with blood–brain barrier used to calculate the membrane permeability. The derivatives showed less efficiency of membrane permeability than control compound. The low aqueous solubility of derivatives may significantly affect its absorption and distribution. Higher doses may be required for bioavailability. All derivatives showed higher lipo affinity than control compound. Intestinal permeability has been found to be comparatively lower than control compound. Derivatives 18 and 25 also have a molecular weight >500.

Toxicity Indicated by Quinazolinone Derivatives at High Doses/Long Term use

If administered in high doses or used therapeutically in long term, the toxicity risk assessment screening results indicated (Table 5). One noticeable component is that no any derivative showed carcinogenicity. It indicates the safe trials of these compounds for further lead optimization. The prior studies related to cases of accumulation and its toxicity also supported the predicted results. Still, there is a scope for further lead optimization based on these calculated ADMET parameters.

Table 4: Lipinski's Rule of Five and Other Parameters for ADME Property Analysis of Molecules

| Name | Lipoaffinity Index | No. of HBA | No. of HBD | LogP | No. of Rotatable bonds | Lipinski Failures | Polar Surface Area | Molecular Weight |
|---------|-----------------------|---------------|---------------|------|------------------------------|----------------------|--------------------------|---------------------|
| Kainate | 1.484339 | 5 | 1 | 2.01 | 4 | 0 | 46.17 | 212.0923 |
| 10 | 7.171926 | 7 | 2 | 2.89 | 4 | 0 | 62.88 | 387.1331 |
| 11 | 11.25937 | 7 | 2 | 3.88 | 5 | 0 | 49.74 | 473.1852 |
| 12 | 8.866538 | 7 | 2 | 3 | 4 | 0 | 49.74 | 465.0759 |
| 13 | 9.062381 | 7 | 2 | 3.22 | 4 | 0 | 49.74 | 445.1306 |
| 14 | 7.70286 | 7 | 2 | 2.89 | 5 | 0 | 95.56 | 490.1156 |
| 15 | 5.079652 | 7 | 2 | 3.44 | 4 | 0 | 49.74 | 470.1384 |
| 16 | 6.531924 | 7 | 2 | 3.11 | 3 | 0 | 49.74 | 432.1227 |
| 17 | 7.257386 | 7 | 2 | 2.78 | 4 | 0 | 62.88 | 421.0942 |
| 18 | 11.34594 | 7 | 2 | 3.77 | 5 | 2 | 49.74 | 507.1462 |
| 19 | 7.355385 | 7 | 2 | 2.78 | 5 | 0 | 95.56 | 476.1 |
| 20 | 7.562317 | 7 | 2 | 3 | 5 | 0 | 95.56 | 456.1546 |
| 21 | 6.215547 | 7 | 2 | 2.67 | 6 | 0 | 141.38 | 501.1397 |
| 22 | 3.545965 | 7 | 2 | 3.22 | 5 | 0 | 95.56 | 481.1624 |
| 23 | 5.019499 | 7 | 2 | 2.89 | 4 | 0 | 95.56 | 443.1468 |
| 24 | 5.777762 | 7 | 2 | 2.56 | 5 | 0 | 108.7 | 432.1182 |
| 25 | 9.802674 | 7 | 2 | 3.55 | 6 | 2 | 95.56 | 518.1703 |
| 5 | 8.778759 | 7 | 2 | 3.11 | 4 | 0 | 49.74 | 431.1149 |
| 6 | 8.975218 | 7 | 2 | 3.33 | 4 | 0 | 49.74 | 411.1695 |
| 7 | 7.623879 | 7 | 2 | 3 | 5 | 0 | 95.56 | 456.1546 |
| 8 | 4.661845 | 7 | 1 | 3.66 | 4 | 0 | 61.42 | 447.1695 |
| 9 | 6.448959 | 7 | 2 | 3.22 | 3 | 0 | 49.74 | 398.1617 |

Complete Molecule evaluation Profile

To evaluate the performance of compounds, four basic strategies: pharmacophore mapping, QSAR, docking & ADMET screening, were implemented. Although predicted Ki was showed mild activity against glutamate receptor, but conclusively, 22, 15 and 8 were observed to be most feasible to act against glutamate receptor in **table 5**.

Final ADMET Compounds Pharmacophore **QSAR** Docking Remark Matched Positive Z-Binding pp-Non-Nono-easy pp-BBB+ ША+ Affinity More Features with Caco+ Score Inhibitor metabolism Carcinogenicity Substrate Considered than Control Control Control Kainate (PDB: 1GR2) ī

Table 5: Complete Molecule Evaluation Profile

CONCLUSIONS

We report here the establishment of 2D-QSAR model and docking study on a series of quinazolinone derivatives as glutamate inhibitors. The performance of quinazolinone derivatives for anticonvulsant activity against glutamate receptor, four basic strategies: pharmacophore mapping, QSAR, docking & ADMET screening were implemented. Although predicted Ki was showed mild activity against glutamate receptor, but conclusively, compounds 22, 15 and 8 were observed to be most feasible to act against glutamate receptor. The correlation of the results obtained from docking and QSAR studies lead to better understanding of the structural requirements for enhanced activity. The obtained results can be used as a guideline to design and predict new potent glutamate inhibitors, which could be an effective way to find novel leads for the development of the anticonvulsant drug.

CONFLICT OF INTEREST

The authors report no conflict of interest.

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REFERENCES

- 1. McNamara JO., 2001. Drugs effective in the therapy of the epilepsies. 2001, In: Goodman and Gillman are the pharmacological basis of therapeutics. Hardman, JG, Limbird LE (eds). 10th Ed. New York, McGraw-Hill, 521-39
- 2. Loscher W., 1998. New visions in the pharmacology of anticonvulsion. Eur J Pharmacol. 19; 342(1):1-13.
- 3. Mattson RH; 1992. Drug treatment of partial epilepsy. Adv Neurol. 57: 643-50.
- 4. Porter RK, Penry JK. in: Meinardi, H, Rowan AJ; 1978; (Eds.), Advances in Epileptology: Psychology, Pharmacotherapy and new Diagnostic Approaches, Amsterdam, Netherlands, 220.
- 5. Gupta YK, Malhotra J; 1997. An adenosinergic system as an endogenous anticonvulsant mechanism. J Physiol Pharmacol. 41, 329-43.
- 6. Coyle JT, Leski M, Morrison JH. (2002). The diverse roles of L-glutamic acid in brain signal transduction. In: Davis KL, Charney D, Coyle JT, Nemeroff C. (Eds.). Neuropsychopharmacology: The Fifth Generation of Progress. Lippincott, Williams, & Wilkins, Philadelphia PA, pp.71–90.
- 7. Brodie MJ; 1990. Status epilepticus in adults. Lancet. 336, 551-2.
- 8. Wolfe JF, Rathman TL, Sleeve MC et al; 1990. Synthesis and anticonvulsant activity of some new 2-substituted 3-aryl-4(3H)-quinazolinones. J. Med. Chem. 33, 161-166.
- 9. Pandolfo, M., 2011. Genetics of epilepsy. Semin Neurol 31, 506-518.
- 10. Berkovic, S. F., Mulley, J. C., Scheffer, I. E., Petrou, S., 2006. Human epilepsies, interaction of genetic and acquired factors. Trends Neurosci, 29, 391-7.
- 11. Rassner, M. P., Moser, A., Follo, M., Joseph, K., van Velthoven-Wurster, V., Feuerstein, T. J., 2016. Neocortical GABA release at high intracellular sodium and low extracellular calcium: an anti-seizure mechanism. J Neurochem 137, 177-89.
- 12. Czapinski, P., Blaszczyk, B., Czuczwar, S. J., 2005. Mechanisms of action of antiepileptic drugs. Curr Top Med Chem 5, 3-14.
- 13. Hanada T, Hashizuma Y, Tokuhara N, et al. Perampanel, (2011). A novel orally active, noncompetitive AMPA-receptor antagonist that reduces seizure activity in rodent models of epilepsy, Epilepsia, 52, 1331-1340.
- 14. Gibbs JW 3rd, Sombati S, De Lorenzo RJ, Coulter DA,(2001). Cellular actions of topiramate, blockade of Kainate –evoked inward currents in cultured hippocampal neurons, Epilepsia, 41(Suppl.1), S10-S16.
- 15. Pandeya S.N., 2012. Semicarbazone- a versatile therapeutic pharmacophore for fragment based anticonvulsant drug design, Acta Pharm. 62, 263-286.
- 16. Ladha SS, Bhatnagar SP; 2008.Rapid microwave-assisted solution phase synthesis of 6,8- disubstituted-2-phenyl-3-(substituted-benzothiazole-2-yl)-4-[3H]-quinazolinone as novel anticonvulsants. Phosphorus, Sulfur, and Silicon and the Related Elements.183(9), 2262-2273
- 17. Global Burden of Disease Study, C., 2015. Global, regional, and national incidence, prevalence, and years lived with disability for 301 acute and chronic diseases and injuries in 188 countries, 1990-2013: a systematic analysis for the Global Burden of Disease Study 2013. Lancet 386, 743-800.
- 18. L.Tripathi and P. Kumar; 2013. Augmentation of GABAergic neurotransmission by novel N-(substituted)-2-[4-(substituted)benzylidene] hydrazine carbothioamides-A potential anticonvulsant approach. Eur. J. Med. Chem., 64, 477-487.

- 19. P. Kumar, B. Shrivastava, S.N. Pandeya, L. Tripathi and J.P. Stables; 2012. Design, synthesis and anticonvulsant evaluation of some novel 1, 3 benzothiazol-2-yl hydrazones/acetohydrazones. Med. Chem. Res., 21, 2428-2442.
- 20. Ajeet, L Tripathi, P. Kumar; 2013. Designing of Novel 6(H)-1,3,4-Thiadiazine Derivatives as MMP12 Inhibitors: An MLR and Docking Approach. Am J Pharmacol Sci, 2, 29-34.
- 21. Trott, O., Olson, A. J., 2010. AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. J Comput Chem 31, 455-61.
- 22. Wang, Y., Xiao, J., Suzek, T. O., Zhang, J., Wang, J., Bryant, S. H., 2009. PubChem: a public information system for analyzing bioactivities of small molecules. Nucleic Acids Res 37, W623-33.
- 23. Yap, C. W., 2011. PaDEL-descriptor: open source software to calculate molecular descriptors and fingerprints. J Comput Chem 32, 1466-74.
- 24. Cheng, F., Li, W., Zhou, Y., Shen, J., Wu, Z., Liu, G., Lee, P. W., Tang, Y., 2012. admetSAR: a comprehensive source and free tool for assessment of chemical ADMET properties. J Chem Inf Model 52, 3099-105.
- 25. C.A. Lipinski, L. Lombardo, B. W. Dominy, P.J. Feeney., 2001. Adv. Drug Deliv. Rev. 46, 3-26.
- 26. Trott, O., Olson, A. J., 2010. AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. J Comput Chem 31, 455-461.